De novo structural ensemble determination from single molecule X-ray scattering



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Abstract

Single molecule X-ray scattering experiments with free electron lasers have opened a new route to the structure determination of biomolecules. Because typically only very few photons per scattering image are recorded, structure refinement is quite challenging. In addition, in each scattering event the orientation of the biomolecule is random and unknown. Further, many biomolecules show structural heterogeneity and conformational transitions between different distinct structures; these structural dynamics are averaged out by existing refinement methods.

To overcome these limitations, here we developed and tested a rigorous Bayesian approach and demonstrate that it should be possible to determine not only a single structure, but an entire structural ensemble from these experiments. Using 10^7 synthetic scattering images generated from molecular dynamics trajectories, our approach was able to resolve the unfolded ensemble of the miniprotein chignolin at 4 - 7 Å resolution. These findings show that X-ray scattering experiments using state-of-the-art free electron lasers should allow one to determine not only biomolecular structures, but whole structure ensembles and, ultimately, 'molecular movies'. In the experiments, a stream of single biomolecules enters a pulsed femtosecond high-intensity X-Ray free electron laser (XFEL) beam, and for each hit the positions of the scattered photons are recorded as an image.

- 10⁶ to 10⁹ images can be recorded with pulse rates of over 20 kHz
- 10-50 photons per image are scattered for typical proteins.
- Images are very noisy. Up to 90% of the scattered photons are incoherent and do not contain structural information.
- Orientations are unknown.
- Conformations are unknown.

Potentially, this allows determination of not only single structures but whole structural ensembles. However, previously available analysis methods¹⁻³ require at least 100-1000 photons per image to determine multiple structures.

Introduction

orientation unknown conformation unknown

single Biomolecules



Almost no experimental data is available yet. Therefore we use synthetic data to test our approach.





A. Our Bayesian approach determines a weighted ensemble of structures by maximizing or sampling from the Bayesian functions. The optimal positions of these Gaussians are posterior probability, computed using Bayes' theorem.
C. The structures are represented as a sum of Gaussian functions. The optimal positions of these Gaussians are determined using simulated annealing.

B. The likelihood decomposes into a product of the **D**. To enhance convergence, sampling is performed in multiple probabilities of the independent images, and for each image, includes a weighted ensemble average and an integral over corresponding to smoothed structures are generated by rejection all possible orientations of the molecule.



with Gaussian

(remove photons)

